Research Article

Gd³⁺-Doped Magnetic Nanoparticles for Biomedical Applications


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Magnetic nanoparticles (MNPs) made of iron oxides with cubic symmetry (Fe₃O₄, γ-Fe₂O₃) are demanded objects for multipurpose in biomedical applications as contrast agents for magnetic resonance imaging, magnetically driven carriers for drug delivery, and heaters in hyperthermia cancer treatment. An optimum balance between the right particle size and good magnetic response can be reached by a selection of a synthesis method and by doping with rare earth elements. Here, we present a microwave-assisted polyol synthesis of iron oxide MNPs with actual gadolinium (III) doping from 0.5 to 5.1 mol.%. The resulting MNPs have an average size of 14 nm with narrow size distribution. Their surface was covered by a glycol layer, which prevents aggregation and improves biocompatibility. The magnetic hyperthermia test was performed on 1 and 2 mg/ml aqueous colloidal solutions of MNPs and demonstrated their ability to rise the temperature by 3 °C during a 20–30 min run. Therefore, the obtained Gd³⁺ MNPs are the promising material for biomedicine.

1. Introduction

Magnetic nanoparticles (MNPs) have been attracting special attention of researchers in recent years due to their unique properties such as size-dependent magnetic behavior (including large magnetic moment and superparamagnetism), low toxicity for living organisms, and possession of active surface for the adsorption/immobilization of ligands and other metals [1]. They have possible applications in catalysis [2, 3], biomedicine [4], water purification from heavy metals [5, 7], magnetic data recording devices [8], and so on. MNPs can be obtained by a variety of wet chemical methods, including a sol–gel method with the use of NaCl and NaH₂PO₄ to control initial aggregation [9], a hydrothermal synthesis with polyethyleneimine as a capping ligand [10], and a solvothermal synthesis in ethylene glycol (EG) with oleylamine and 1,3-diaminopropane as surfactants and sodium acetate as a steric stabilizer [11]. In particular, the use of glycols in the synthesis allows producing MNPs with increased hydrophilicity and improved biocompatibility.

Among magnetic materials for nano-biomedicine, magnetite (Fe₃O₄) and maghemite (γ-Fe₂O₃) are the most widely used. Both oxides are ferrimagnetic; however, in a nanconfined form, they exhibit superparamagnetic behavior, where magnetic susceptibility is higher than that in paramagnetic materials [12]. Magnetite is a preferred material for biomedical applications due to a larger magnetic moment and availability of relatively simple synthesis methods. It has been reportedly used in magnetic cell separation [13], magnetocytolysis [14], magnetic hyperthermia [15], tracking of
2. Materials and Methods

2.1. Synthesis of MNPs. The reagents were purchased from commercial suppliers (Alfa Aesar and Sigma-Aldrich) and used as such. The initial molar fraction of gadolinium ions in respect to total amount of iron and gadolinium ions was varied from 0.5 to 9 mol.%. In a typical synthesis (Table 1 for the whole set of samples), 0.665 g of anhydrous iron (III) chloride and 0.001 g of gadolinium (III) chloride were dissolved in 10 ml of triethylene glycol. A solution of 0.096 g of sodium hydroxide in 7.4 ml of triethylene glycol was prepared apart. All solutions were mixed at room temperature and then continuously stirred under flowing argon for 30 min. The reaction mixture was placed in a MW reactor CEM Discover SP (175 W) and kept at 220°C for 1.5 h with a medium rate stirring. The fine black-colored colloidal MNPs were formed in the reaction mixture. The mixture was cooled to room temperature, and the particles were collected with a permanent magnet. They were washed 5 times with distilled water and once with ethyl alcohol and then dried in vacuum at 40°C for 12 h. The samples were labeled according to the actual gadolinium content as shown in Table 1.

2.2. Characterization Techniques. Samples were examined on a Bruker D2 Phaser X-ray diffractometer (Cu Kα, λ = 1.5417 Å) with a step of 0.02°. X-ray diffraction (XRD) patterns were analyzed by Jana2006 program [27], following the procedure elsewhere [28, 29]. X-ray fluorescence (XRF) analysis was carried out on a two-dimensional micro-X-ray fluorescence spectrometer Bruker M4 Tornado. At least 400 particles were analyzed for every sample to build a particle size distribution histogram. High-resolution TEM (HR-TEM) and scanning TEM (STEM), selected area electron diffraction (SAED), and energy dispersive spectroscopy (EDS) imaging were performed on FEI Tecnai G2 Spirit BioTWIN. At least 400 particles were analyzed for every sample to build a particle size distribution histogram. High-resolution TEM (HR-TEM) and scanning TEM (STEM), selected area electron diffraction (SAED), and energy dispersive spectroscopy (EDS) imaging were performed on FEI Tecnai G2 F20S-TWIN. Infrared (IR) spectra were measured on a Fourier-transform infrared (FTIR) Bruker Vertex 70 spectrometer in transmittance with 64 scans per spectrum of 2 cm⁻¹ resolution, pelleting the samples with KBr in 1 : 150 weight ratio. Spectra were converted in the absorbance values and baseline corrected (by a standard rubberband correction) using the instrument’s Opus software. Calorimetric measurements for calculation of specific absorption rate (SAR) were done on a LocalHyperThermLabUnit TOR 04/16 experimental setup (Nanomaterials LLC, Russia) at 218.9 kHz with a magnetic field flux of 20 mT (15923 Oe).

3. Results and Discussion

3.1. Crystalline Phase and Actual Gd³⁺ Content of MNPs. Since gadolinium doping of iron oxides proceeds inefficiently due to a large difference in ionic radii of Gd³⁺ and Fe³⁺, the first thing to control after the synthesis was the actual quantity of the dopant in the samples. The values were processed from XRF data and presented in Table 1 together with respective initial loading of gadolinium. As it comes from comparison of Table 1, the actual content of Gd³⁺ ions in the resulting material is significantly lower than the initially loaded one. This is an undesirable fact, but similar situation was already observed by other authors reporting on
iron oxides doping with rare earth elements [18]. Let us briefly discuss the possible reasons without entering into matters of detail. The aforementioned difficulty for the larger Gd\(^{3+}\) ion to accommodate in the iron oxide lattice implies a fraction of gadolinium being preserved in the reaction mixture in either a form of unreacted salt or, more likely, as a hydroxide (Gd(OH)\(_3\)). At the end of synthesis, it would not be recovered with a magnet and will not remain in the final material. Since Gd\(^{3+}\) ion preferentially occupies the octahedral sites of the inverse spinel structure of \(\text{Fe}_3\text{O}_4\) while hardly entering the smaller tetrahedral sites, only a certain relatively high concentration of gadolinium will be needed to set the conditions for its incorporation into the crystal lattice in a greater extent.

Figure 1 presents the XRD patterns of the synthesized iron oxides with a different Gd\(^{3+}\) content as well as that of pristine iron oxide from the same synthesis (patterns are vertically shifted for clarity). Most of the patterns (excluding those for the samples with gadolinium doping of 2.5 and 4.3 mol.%) contain the same sequence of the reflections at \(2\theta = 30.2, 35.5, 43.2, 53.5, 57.2, \) and 62.7° indicative for an inverse spinel phase and corresponding to (220), (311), (400), (422), (511), and (440) crystallographic planes, respectively, which is immanent to both magnetite and maghemite [1]. There are no other reflections in these patterns, which evidences the phase purity of the samples. This would mean that gadolinium is evenly dispersed in the lattice without forming a distinct phase. Both MNP-Gd2.5% and MNP-Gd4.3% samples contain an additional orthorhombic phase, which was assigned to goethite (α-FeOOH). The established pure phases and their mixtures are summarized in Table 1 in terms of cubic and orthorhombic lattice systems.

We noted that formation of the iron hydroxide phase occurs at relatively high Gd\(^{3+}\) loading. This might be a result of excessive presence of the Fe\(^{3+}\) ions in the reaction mixture due to favorable incorporation of Gd\(^{3+}\) ions into the growing particles. We will illustrate this assumption by equations of chemical reactions taking place during the synthesis of MNPs.

![Figure 1: XRD patterns of synthesized iron oxides with different Gd\(^{3+}\) content.](image)

**Figure 1:** XRD patterns of (1) MNP-Gd0%, (2) MNP-Gd0.5%, (3) MNP-Gd0.6%, (4) MNP-Gd2.5%, (5) MNP-Gd4.3%, and (6) MNP-Gd5.1%. The solid dots indicate the diffractions of the pure magnetite phase (JCPDS 01-074-1910).

The magnetite MNPs are formed in a coprecipitation reaction:

\[2\text{Fe}^{3+} + \text{Fe}^{2+} + 8\text{OH}^- \rightarrow \text{Fe}_3\text{O}_4 + 4\text{H}_2\text{O} \quad (1)\]

Presence of the Fe\(^{2+}\) ions can be explained by the partial reduction of the Fe\(^{3+}\) ions from the iron precursor according to the following reactions [24]:

\[\text{CH}_3\text{OH} + \text{CH}_2\text{OH} \rightarrow \text{CH}_3\text{CHO} + \text{H}_2\text{O} \quad (2)\]

\[2\text{Fe}^{3+} + 2\text{CH}_3\text{CHO} + \text{OH}^- \rightarrow 2\text{Fe}^{2+} + \text{CH}_3\text{CO} - \text{COCH}_3 + 2\text{H}_2\text{O} \quad (3)\]

The excess of Fe\(^{3+}\) ions (that does not participate in formation of MNPs due to the presence of competitive Gd\(^{3+}\) ions) can react with abundant hydroxide ions:

![Table 1: Properties of synthesized MNPs.](image)

**Table 1:** Properties of synthesized MNPs: initial loading (\(\chi_{\text{ini}}\)) and actual content (\(\chi_{\text{act}}\) by XRF data) of Gd\(^{3+}\) ions in the samples, a cubic lattice parameter (\(a\)), and average crystallite size of MNPs (\(d_{av}\)) calculated from XRD and TEM data, and SAR values evaluated after the hyperthermia test.

<table>
<thead>
<tr>
<th>Sample name</th>
<th>(\chi_{\text{ini}}) Gd(^{3+}) (mol.%)</th>
<th>(\chi_{\text{act}}) Gd(^{3+}) (mol.%)</th>
<th>Lattice system</th>
<th>(a) (XRD) (Å)</th>
<th>(d_{av}) (XRD) (nm)</th>
<th>(d_{av}) (TEM) (nm)</th>
<th>SAR (W/g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MNP-Gd0%</td>
<td>0</td>
<td>0</td>
<td>Cubic(^4)</td>
<td>8.368(8)</td>
<td>6</td>
<td>6.6</td>
<td>15.6</td>
</tr>
<tr>
<td>MNP-Gd0.5%</td>
<td>1</td>
<td>0.5</td>
<td>Cubic(^4)</td>
<td>8.385(2)</td>
<td>10</td>
<td>14.9</td>
<td>12.5</td>
</tr>
<tr>
<td>MNP-Gd0.6%</td>
<td>2.7</td>
<td>0.6</td>
<td>Cubic(^4)</td>
<td>8.348(3)</td>
<td>15</td>
<td>15.5</td>
<td>15.9</td>
</tr>
<tr>
<td>MNP-Gd2.5%</td>
<td>5.6</td>
<td>2.5</td>
<td>Cubic + orth.(^5)</td>
<td>8.366(5)</td>
<td>14</td>
<td>14.1</td>
<td>15.5</td>
</tr>
<tr>
<td>MNP-Gd5.1%</td>
<td>7</td>
<td>5.1</td>
<td>Cubic(^4)</td>
<td>8.386(1)</td>
<td>15</td>
<td>13.3</td>
<td>18.0</td>
</tr>
<tr>
<td>MNP-Gd4.3%</td>
<td>9</td>
<td>4.3</td>
<td>Cubic + orth.(^5)</td>
<td>—</td>
<td>—</td>
<td>8.5</td>
<td>13.1</td>
</tr>
</tbody>
</table>

\(^4\)Pure inverse spinel phase (\(\text{Fe}_3\text{O}_4\) or/and γ-\(\text{Fe}_2\text{O}_3\)); \(^5\)orthorhombic goethite (α-FeOOH with \(a = 4.59(1)\) Å, \(b = 9.90(5)\) Å, and \(c = 3.00(1)\) Å) and spinel phase; \(^\ddagger\)orthorhombic goethite (α-FeOOH with \(a = 4.590(3)\) Å, \(b = 9.935(8)\) Å, and \(c = 3.025(2)\) Å) and spinel phase.
Fe$^{3+} + 3\text{OH}^- \rightarrow \text{Fe(OH)}_3 \quad (4)

Then, iron (III) hydroxide can form goethite by partial dehydration [30]:

$$\text{Fe(OH)}_3 \rightarrow \text{FeO(OH)} + \text{H}_2\text{O} \quad (5)$$

It is unlikely for the Gd$^{3+}$ ions to enter the growing crystalline lattice of the goethite phase. Such sample will combine NPs of two phases, what we have actually seen from XRD data. This also explains why the lower gadolinium content was detected in the resulting sample after initial loading of 9 mol.% compared to that of 7 mol.%

For the MNPs of a spinel phase, we have calculated the lattice parameter and the average crystallite size (by Debye–Scherrer equation). The respective values are presented in Table 1.

3.2. MNPs Size, Morphology, and Elements Distribution. The results of TEM measurements are presented in Figure 2 together with respective particle size distribution histograms. All Gd$^{3+}$-doped samples (except MNP-Gd5.1%) consist of both spherical (smaller) and polyhedral (bigger) particles with sizes in the range from 6 to 24 nm. Particle size distribution is narrow, and its profile is close to normal one for the most cases. Both MNP-Gd4.3% and MNP-Gd0% samples are characterized by more tight distribution interval (from 6 to 10 nm). The weighted average values of MNPs size for each sample were calculated from the obtained distributions and presented in Table 1. It can be seen that regardless of the gadolinium content the average size of MNPs is 14 nm (excluding the MNP-Gd4.3% sample). This observation implies that the adopted synthesis method shows a good reproducibility in terms of particle size, and its parameters (such as time, duration, and concentration of metal precursors in reaction volume) would define the main dimension of the resulting MNPs.

The sample with relatively high gadolinium content, MNP-Gd5.1%, was chosen for the studies by applying the advanced electron microscopy techniques (Figure 2). The HR-TEM images (Figures 3(a) and 3(b)) confirmed a polyhedral shape of MNPs. The obtained SAED images (Figure 3(c)) allow calculating the values of interplanar distances as 2.92 and 2.52 Å, which correspond to (311) and (220) planes of the Fe$_3$O$_4$ cubic phase, respectively. The measured interplanar spacing is 0.294 nm, evidencing the (220) plane of the cubic structure of magnetite. The combined STEM-EDS mapping (Figures 3(d) and 3(e)) on the same region of the sample evidences that iron and gadolinium are uniformly distributed across the volume of MNPs. As presented at the EDS histogram (Figure 3(f)), the total weight content of gadolinium and iron in a given point of the MNP-Gd5.1% sample can be estimated as 6.8 and 71.86%, respectively. Such weight ratio corresponds to 3.3 mol.% of gadolinium. This value is 1.5 times lower than that estimated from the XRF data. Such difference is often happening when two dissimilar techniques are used.

3.3. MNPs Vibrational Profile and Surface Coverage. FTIR spectra of the synthesized samples are shown in Figure 4(a) (vertically shifted for clarity). The dominating complex band below 700 cm$^{-1}$ is due to MNPs stretching modes. At first sight, its complex profile does not change appreciably with increasing Gd content. Instead, when normalized at maximum intensity (Figure 4(b)), the band’s shape changes with gadolinium content, most notably at about 630 cm$^{-1}$. The profile of this band can be fitted with tree lorentzians (Figure 4(b)) with maxima falling at 571, 630, and 700 cm$^{-1}$. The first frequency is a typical γFe–O vibration in magnetite [31]. That is one of the two characteristic bands of the Fe$_3$O$_4$ phase at 570 cm$^{-1}$ ($\nu_1$) and at 390 cm$^{-1}$ ($\nu_2$) [32]. They originate from the Fe–O stretching mode of the tetrahedral and octahedral sites for the $\gamma_1$ frequency band and the Fe–O stretching mode of the octahedral sites for the $\gamma_2$ frequency band. Introduction of defects into the structure causes widening and eventual splitting of both $\gamma_1$ and $\gamma_2$ bands, giving the rise to new IR bands at around 430 and 630 cm$^{-1}$. The γ-Fe$_3$O$_4$ phase is considered as a defective form of Fe$_3$O$_4$ and it has characteristic bands at 700 and 630 cm$^{-1}$ [33]. Turning to our samples, we found that the component at 630 cm$^{-1}$ exhibits a slight growth with increasing gadolinium content (Figure 4(b)), supporting an assumption about introduction of Gd$^{3+}$ ions into the Fe$_3$O$_4$ sublattice of the Fe$_3$O$_4$ crystal phase of MNPs. Similar trend can be found when considering evaluation of the shape in a complimentary three-component band at lower frequencies. For the sake of clarity, we report its enlarged profile and fitting results for just two samples of MNP-Gd0% and MNP-Gd5.1% (Figure 4(c)). By comparing the relative intensities of fitting components, the rise of the band at 440 cm$^{-1}$ with increasing gadolinium content is clearly evident.

Another iron compound, whose presence has been assumed in the samples based on the XRD data, is goethite, which expresses the –OH stretch vibrations at 1124, 890, and 810 cm$^{-1}$ [33]. These frequencies comply well with the bands clearly visible for the MNP-Gd2.5% and MNP-Gd4.3% samples.

The spectroscopic features in the adjacent 1600–800 cm$^{-1}$ interval contain a vibrational signature of glycol [34] which was involved in the synthesis and was preserved in the samples. Its spectral fingerprint is easily seen in spectrum of the MNP-d2.5% sample. In brief, two groups of bands at about 1100 and 900 cm$^{-1}$ originate from out-of-plane and in-plane γCH$_2$ vibrations (bands at 1118 and 935 cm$^{-1}$) and stretching modes of skeleton C–O bonds (bands at 1070 and 890 cm$^{-1}$), respectively. A spectral envelope at about 1400 cm$^{-1}$ is composed of various bending modes of CH$_2$ groups.

The distinct band at 1600 cm$^{-1}$ (see MNP-Gd0% in Figure 4) appears after the functionalization of glycol onto the surface of magnetite NPs and will be commented in more detail. The high-frequency side of the band is formed by contribution from the δOH vibration of water moieties falling at ~1630 cm$^{-1}$. The right side of the band contains the strong γC–O vibration, which appears due to a covalent bonding between iron and carbonyl oxygen of glycol [35].
Taking into account a featureless neighboring band at 1400 cm\(^{-1}\), we may assume of being in presence of carboxyl’s oxygen bonding in unidentate arrangement \[13\] due to a large separation (\(\Delta \sim 200\) cm\(^{-1}\)) between the asymmetric (COO\(\equiv\)) and symmetric (COO\(\equiv\)) vibrations. With increasing amount of added Gd\(^{3+}\) ions, this separation decreases (\(\Delta = 135\) cm\(^{-1}\) for the MNP-Gd2.5% sample), evidencing a chelating arrangement of the glycol ligands on the surface of MNPs.

Although stretching modes of hydroxyl groups in glycol fall in the 3700–3600 cm\(^{-1}\) interval, they are hidden by stretching vibrations of water molecules physisorbed on potassium bromide (used for the pelleting), leading to formation of a very broadband in the 3600–3000 cm\(^{-1}\) range. Nearby, there is a complex band at about 2850 cm\(^{-1}\) due to the overlapping asymmetric and symmetric stretching vibrations of methylene groups.

3.4. Hyperthermia Test with MNPs. As it is known, magnetic induction hyperthermia is the treatment of malignant tumors. MNPs embedded around a tumor site and placed under the action of an external oscillating magnetic field will heat up the local environment to a temperature of 41–46 °C, thus killing cancer cells \[36\]. For this treatment, magnetic fluids, that is, polydisperse colloidal solutions of single-domain MNPs (grains), are used. In aqueous system, MNPs tend to aggregate due to the long range Van der Waals’ forces. Presence of a glycol layer on the surface of synthesized samples is beneficial for enhancing the dispersion and stabilizing the suspension.

The hyperthermia measurement was conducted for all samples simultaneously. Before presenting the results, some relevant theoretical considerations have to be mentioned. In an experiment performed over a long heating, the non-adiabatic processes due to heat loss released from the sample to surrounding environment should be taken into account. Such process could be described via change of temperature, \(T\), over time, \(t\) \[37\]:

\[
T(t) = T_0 + \Delta T_{\text{max}} \left(1 - e^{-t/\tau}\right),
\]

where \(T_0\) is the initial temperature (thermostat temperature) of the sample, \(\Delta T_{\text{max}}\) is the maximum change of the temperature in the stable state, and \(\tau\) is the heating time constant.

SAR value can be calculated by the initial slope method, that is, by approximating the slope of the initial heat rise, \(\beta\):

\[
\beta = \frac{dT}{dt}\bigg|_{t=0}.
\]

The underlying assumption is that the initial slope of the curve does not suffer from losses since the sample is at its initial temperature. Therefore, the SAR can be calculated from the following equation:

\[
\text{SAR} = \frac{C m_w}{m_{\text{sample}}} \beta,
\]

where \(C\) is the heat capacity of the sample, \(m_w\) is the mass of the solvent (water), and \(m_{\text{sample}}\) is the total mass of the sample in water \[38\].

The experimental results of the measured time changes in the temperature of magnetic fluids of 1 mg/ml and 2 mg/ml
concentrations under the action of the external magnetic field are presented in Figure 5, while calculated SAR values are summarized in Table 1. The results show that the synthesized materials demonstrate a good response to the oscillating magnetic field. Within 20–30 min of action, the water solution warms up by 3°C. In case of performing hyperthermia on these fluids in vivo, this would allow to reach about 40–41°C, a therapeutically desired level of temperature impact. There is a direct correlation between the concentration of MNPs in the probe and the final heating temperature. The obtained results are comparable with those described in [19], where MNPs having a ten times lower gadolinium content (from 0.02 to 0.08 mol.%) were investigated.

4. Conclusions
The proposed MW-assisted polyol synthesis allows preparing Gd³⁺-doped MNPs in the 6–24 nm size range (TEM) with an average crystallite size of 14 nm (XRD). The actual values of Gd³⁺ doping of materials were in the range of 0.5–5.1 mol.% (XRF). Dopant is uniformly distributed in the volume of MNPs (STEM-EDS). The surface of NPs is
covered by glycol molecules (FTIR). We also found that introduction of gadolinium (III) in quantities higher than 2.7 mol.% alters the purity of a spinel phase of resulting MNPs. Hyperthermia test of 1 and 2 mg/ml aqueous colloidal solutions of MNPs demonstrated their heating ability at 3°C in 20–30 min run. These results confirm the relevance of synthesized materials for biomedical applications.

**Data Availability**

X-ray powder diffraction raw data are available in Supplementary Materials.

**Conflicts of Interest**

The authors declare that there are no conflicts of interest regarding the publication of this paper.

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**Supplementary Materials**

Supplementary materials contain X-ray diffraction patterns in the XYE format, where the first column represents the 2θ values, second column is the scattered intensity, and the final column contains absolute error according to the Poisson statistics. (Supplementary Materials)

**References**


